

Choline as a tool to evaluate nicotinic receptor function in chromaffin cells.

*Juana M^a González-Rubio^a, Jonathan Rojo^a, Laura Tapia^a,
Victoria Maneu^d, José Mule^f, Luis M. Valor^c, Manuel Criado^c,
Francisco Sala^c, Antonio G. García^{a,b} and Luis Gandía^a*

*a. Instituto Teófilo Hernando, Departamento de Farmacología y Terapéutica,
Facultad de Medicina, Universidad Autónoma de Madrid,*

*b. Servicio de Farmacología-Unidad Clínica and Instituto de Gerontología,
Hospital de la Princesa, Madrid*

*c. Instituto de Neurociencias, Universidad Miguel Hernández-CSIC, Sant Joan
d'Alacant,*

*d. Departamento Interuniversitario de Óptica. Universidad de Alicante. San
Vicente del Raspeig, Alicante, Spain*

Correspondence: Dr. Luis Gandía, Instituto Teófilo Hernando.
Dpto. de Farmacología y Terapéutica. Facultad de Medicina.
Universidad Autónoma de Madrid. Arzobispo Morcillo 4, 28029 Madrid, Spain.
Phone: 34-91-4975396; **Fax:** 34-91-4975380; **Email:** luis.gandia@uam.es

Cell Biology of the Chromaffin Cell
R. Borges & L. Gandía Eds.
Instituto Teófilo Hernando, Spain, 2004

Choline is present in the synaptic cleft as a result of the rapid degradation of acetylcholine (ACh) by acetylcholinesterase (AChE). The hypothesis that choline will exert some modulatory function of synaptic neurotransmission is supported by few studies; for instance, choline has muscarinic effects on central neurons¹, evokes catecholamine release by acting as a partial agonist at neuronal nicotinic receptors for ACh (nAChRs), and blocks partially the release of catecholamines evoked by ACh, in cultures of bovine adrenal medullary chromaffin cells². Choline has been also shown as a selective agonist of $\alpha 7$ nAChRs in brain neurons³. As far as we know, a detailed study of the effects of choline on the kinetic properties of inward currents through $\alpha 7$ and $\alpha 3\alpha 4$ nAChRs is not available. Hence, we decided to express homomeric $\alpha 7$ and heteromeric $\alpha 3\alpha 4$ bovine nicotinic receptors in *Xenopus* oocytes and study the effects of choline on these receptors. We report here the results of such study that might be relevant in the frame of the growing number of physiological and physiopathological roles that are being ascribed to brain nAChRs, i.e. in neuroprotection, analgesia, addiction to nicotine, dementia or behaviour^{4,5}.

RESULTS AND DISCUSSION

Application of brief (1-5 s) pulses of choline (0.3-10 mM) elicited inward current in oocytes expressing $\alpha 7$ nAChRs, acting as a full agonist of these receptors (Figure 1) with an EC_{50} of about 0.6 mM. In contrast, in oocytes expressing $\alpha 3\alpha 4$ nAChRs, choline did not elicit any inward current by itself. However, choline blocked the inward current through these receptors, induced by 5-s pulses of 0.1 mM ACh (I_{ACh}) (figure 2), with an IC_{50} of about 0.38 mM. This blockade was quickly reversible after choline washout. The nature of I_{ACh} blockade by choline was further tested by challenging the oocytes with increasing concentrations of ACh (from 10 μ M to 3 mM). In these experiments, $\alpha 3\alpha 4$ oocytes were sequentially stimulated with 5-s pulses of increasing concentrations of ACh, given first in the absence and subsequently in the presence of 1 mM choline. The blockade induced by choline was around 65-70% in all cases, suggesting a non-competitive nature of the blockade, as the case is for hexamethonium⁶.

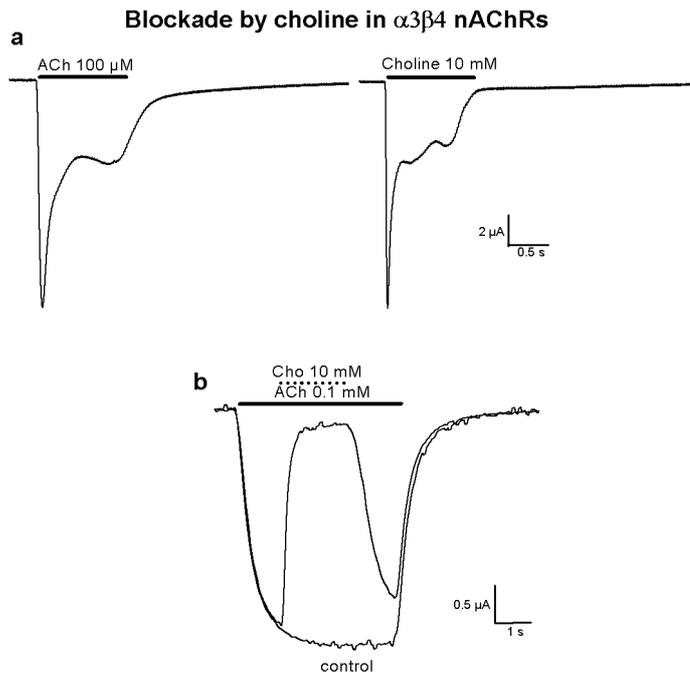


Figure 1. Effects of choline on $\alpha 7$ and $\alpha 3\beta 4$ nAChRs expressed in xenopus oocytes. **a)** Application of choline (10 mM) elicited an inward current in $\alpha 7$ nAChRs, acting as a full agonist of this receptor. In the same oocyte, ACh 0.1 mM elicited similar inward current. **b)** In an oocyte expressing $\alpha 3\beta 4$ nAChRs, ACh 0.1 mM is capable to elicit a response (control) that was fully blocked by choline 10 mM. The blockade of the current was partially recovered when choline was removed from the perfusion solution.

We also wanted to study if the blockade induced by choline of I_{ACh} exhibited voltage-dependence, as has been described for different blockers of nAChRs⁷⁻⁹. For it, a voltage ramp protocol (-100 mV to +60 mV) was applied to $\alpha 3\beta 4$ oocytes voltage-clamped at -80 mV and pulses of ACh (0.1 mM, 5-s) were applied either in the absence or in the presence of increasing concentrations of choline. The I-V curves for the traces are plotted, and calculations were made to estimate the degree of current blockade achieved by each concentration of choline at different voltages. That revealed that I_{ACh} blockade by choline was more pronounced at more hyperpolarised

potentials, as compared to depolarised potentials, in all of the three concentrations of choline tested.

Ours results reported here shown that, with a similar concentration range, choline exerted dual opposite effects in the two main receptor subtypes of nAChRs expressed by bovine chromaffin cells, $\alpha 7$ and $\alpha 3\beta 4$ ¹⁰⁻¹².

The blockade of I_{ACh} through $\alpha 3\beta 4$ receptors was voltage-dependent and was of a non-competitive nature, suggesting that choline had a binding site different to that for the agonist ACh, i.e. choline may bind to the ACh locus on the $\alpha 7$ receptor to cause its activation and to a site different to that of ACh on the $\alpha 3\beta 4$ receptor to cause its blockade. The similar concentration at which choline exerts its dual action on $\alpha 7$ receptor activation and $\alpha 3\beta 4$ receptor blockade, reinforces the hypothesis that choline has a role beyond that of being the simple precursor of ACh.

ACKNOWLEDGMENTS

Supported by research grants from the Ministry of Science and Technology of Spain (N° PM99-004 to LG, BFI2003-02722 to AGG, BMC2002-00972 to MC and PM98-0097 to FS), Programa Grupos Estratégicos III PRICIT de la Comunidad de Madrid, FIS (N° 01/183), Generalitat Valenciana (CTIDIB/2002/138 and GRUPOS03/038), Fundación Teófilo Hernando and Fundación La Caixa.

REFERENCES

1. Krnjevic, K. and W. Reinhardt, *Choline excites cortical neurons*. Science, 1979. **206**:1321-1323.
2. Holz, R.W. and R.A. Senter, *Choline stimulates nicotinic receptors on adrenal medullary chromaffin cells to induce catecholamine secretion*. Science, 1981. **214**:466-468.
3. Alkondon, M., et al., *Choline is a selective agonist of $\alpha 7$ nicotinic acetylcholine receptors in the rat brain neurons*. Eur J Neurosci, 1997. **9**:2734-2742.
4. Jones, S. S. Sudweeks, and J.L. Yakel, *Nicotinic receptors in the brain: correlating physiology with function*. Trends Neurosci, 1999. **22**:555-561.
5. Paterson, D. and A. Nordberg, *Neuronal nicotinic receptors in the human brain*. Prog Neurobiol, 2000. **61**:75-111.
6. Bertrand, D., S. Bertrand and M. Ballivet, *Pharmacological properties of the homomeric alpha7 receptor*. Neurosci Lett, 1992. **146**:87-90.
7. Ascher, P., et al., *Studies on the mechanism of action of acetylcholine antagonist on rat parasympathetic ganglia cells*. J Physiol, 1979. **295**:139-170.

8. Gurney, A.M. and H.P. Rang, *The channel blocking action of methonium compounds on rat submandibular ganglion cells*. Br J Pharmacol, 1984. **82**:623-642.
9. Buisson, B. and D. Bertrand, *Open-channel blockers at the human $\alpha 4\beta 2$ neuronal nicotinic acetylcholine receptor*. Mol Pharmacol, 1998. **53**:555-563.
10. Criado, M., L. Álamo, and A. Navarro, *Primary structure of an agonist binding subunit of the nicotinic acetylcholine receptor from bovine adrenal chromaffin cells*. Neurochem Res, 1992. **17**:281-287.
11. García-Guzmán, M., et al., *Alpha-bungarotoxin-sensitive nicotinic receptors on bovine chromaffin cells: molecular cloning, functional expression and alternative splicing of the alpha 7 subunit*. Eur J Neurosci, 1995. **7**:647-655.
12. Campos-Caro, A., et al., *Neuronal nicotinic acetylcholine receptors on bovine chromaffin cells: cloning, expression, and genomic organization of receptor subunits*. J Neurochem, 1997. **68**:488-497.

